

# Instructions for Use

## Life Science Kits & Assays



**innuPREP Food DNA Kit - IPC16**

**Order No.:**

845-IPS-5716016 16 reactions  
845-IPP-5716016 16 reactions  
845-IPS-5716096 96 reactions  
845-IPP-5716096 96 reactions  
845-IPP-5716480 480 reactions

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This documentation describes the state at the time of publishing.  
It needs not necessarily agree with future versions. Subject to change!

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Made in Germany!      info.innu@ist-ag.com

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# 1 Introduction

## 1.1 Intended use

The innuPREP Food DNA Kit - IPC16 has been designed for automated isolation of DNA from food samples using the InnuPure C16 / C16 *touch*. The extraction procedure is based on a new-patented chemistry.

The extraction procedure starts with an external lysis step of food samples. After the lysis step the sample is transferred into the Reagent Strips or Reagent Plate of the kit, which is already prefilled with all extraction reagents needed for the extraction process. The following extraction process runs automatically on the InnuPure C16 / C16 *touch*. The extraction process is based on binding of the DNA on surface modified magnetic particles. After washing steps the nucleic acid is eluted from the magnetic particle with RNase-free water and is now ready to use for downstream applications. The extraction chemistry in combination with the InnuPure C16 / C16 *touch* protocol are optimized to get maximum of yield and quality.

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### CONSULT INSTRUCTION FOR USE











This package insert must be read carefully before use. Package insert instructions must be followed accordingly. Reliability of results cannot be guaranteed if there are any deviations from the instructions in this package insert.

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## 1.2 Notes on the use of this manual and the kit

For easy reference and orientation, the manual and labels use the following warning and information symbols as well as the shown methodology:

Symbol	Information
	<b>REF</b> Catalogue number.
	<b>Content</b> Contains sufficient reagents for <N> tests.
	<b>Storage conditions</b> Store at room temperature, unless otherwise specified.
	<b>Consult instructions for use</b> This information must be observed to avoid improper use of the kit and the kit components.
	<b>Expiry date</b>
	<b>Lot number</b> The number of the kit charge.
	<b>Manufactured by</b> Contact information of manufacturer.
	<b>For single use only</b> Do not use components for a second time.
	<b>Note / Attention</b> Observe the notes marked in this way to ensure correct function of the device and to avoid operating errors for obtaining correct results.

The following systematic approach is introduced in the manual:

- The chapters and figures are numbered consecutively.
- A cross reference is indicated with an arrow (e.g. → “Notes on the use of this manual” p. 3).
- Working steps are numbered.

## 2 Safety precautions

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### NOTE

Read through this chapter carefully before use to guarantee your own safety and a trouble-free operation.

Follow all the safety instructions explained in the manual, as well as all messages and information, which are shown.

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All due care and attention should be exercised in handling the materials and reagents contained in the kit. Always wear gloves while handling these reagents and avoid any skin contact! In case of contact, flush eyes or skin with a large amount of water immediately.

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### FOR SINGLE USE ONLY!

This kit is made for single use only!

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### ATTENTION!

Don't eat or drink components of the kit!

The kit is designed to be handled by educated personnel in a laboratory environment!

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If the buffer bottles are damaged or leaking, wear gloves and protective goggles when discarding the bottles in order to avoid any injuries. This kit is to be used with potential infectious human samples. Therefore, all liquid waste must be considered as potentially infectious and must be handled and discarded according to local safety regulation.

Please observe the federal, state and local safety and environmental regulations. Follow the usual precautions for applications using extracted nucleic acids. All materials and reagents used for DNA or RNA isolation should be free of DNases or RNases.

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### ATTENTION!

Do not add bleach or acidic components to the waste after sample preparation!

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**NOTE**

Emergency medical information in English and German can be obtained 24 hours a day from:

Poison Information Center, Freiburg / Germany

Phone: +49 (0)761 19 240.

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For more information on GHS classification and the Safety Data Sheet (SDS) please contact [sds.innu@ist-ag.com](mailto:sds.innu@ist-ag.com).

### 3 Storage conditions

The kit is shipped at ambient temperature.

Upon arrival store lyophilized and dissolved **Proteinase K** at 4 °C to 8 °C.

All other components of the innuPREP Food DNA Kit – IPC16 should be stored dry at room temperature (15 °C to 30 °C). When stored at room temperature, the kit is stable until the expiration date printed on the label on the kit box.

Before every use make sure that all components have room temperature. If there are any precipitates within the provided solutions dissolve these precipitates by careful warming.

## 4 Functional testing and technical assistance

The IST Innuscreen GmbH guarantees the correct function of the kit for applications as described in the manual. This product has been produced and tested in an ISO 13485 certified facility.

We reserve the right to change or modify our products to enhance their performance and design. If you have any questions or problems regarding any aspects of the innuPREP Food DNA Kit – IPC16 or other IST Innuscreen GmbH products, please do not hesitate to contact us.

For technical support or further information in Germany please dial +49 30 9489 3380. For other countries please contact your local distributor.

## 5 Product use and warranty

The kit is not designed for the usage of other starting materials or other amounts of starting materials than those, referred to in the manual (→ "Product specifications" p. 8 ). Since the performance characteristics of IST Innuscreen GmbH kits have just been validated for the application described above, the user is responsible for the validation of the performance of IST Innuscreen GmbH kits using other protocols than those described below. IST Innuscreen GmbH kits may be used in clinical diagnostic laboratory systems after the laboratory has validated the complete diagnostic system as required by CLIA' 88 regulations in the U.S. or equivalent regulations required in other countries.

All products sold by the IST Innuscreen GmbH are subjected to extensive quality control procedures and are warranted to perform as described when used correctly. Any problems should be reported immediately.



## 6 Kit components

### 6.1 Included kit components

	$\Sigma$ 16	$\Sigma$ 96	$\Sigma$ 480
<b>REF</b>	845-IPS-5716016 <sup>a</sup> 845-IPP-5716016 <sup>b</sup>	845-IPS-5716096 <sup>a</sup> 845-IPP-5716096 <sup>b</sup>	845-IPP-5716480 <sup>b</sup>
Lysis Solution CBV	25 ml	2x 120 ml	3x 250 ml
Proteinase K	for 2 x 0.3 ml working solution	for 2 x 1.5 ml working solution	for 7 x 1.5 ml working solution
Reagent Strip M <sup>a</sup>	16 (pre-filled, sealed)	96 (pre-filled, sealed)	--
Reagent Plate M <sup>b</sup>	2 (pre-filled, sealed)	12 (pre-filled, sealed)	60 (pre-filled, sealed)
Filter Tips	2 x 16	2 x 96	10 x 96
Elution Tubes (0.65 ml)	16	2 x 48	10 x 48
Elution Caps (Stripes)	2	12	5 x 12
Elution Stripes	2	12	5 x 12
Manual	1	1	1
Initial steps	<b>Proteinase K</b> Dissolve by addition of 0.3 ml of ddH <sub>2</sub> O, mix thoroughly and store as described above.	<b>Proteinase K</b> Dissolve by addition of 1.5 ml of ddH <sub>2</sub> O, mix thoroughly and store as described above.	

### 6.2 Components not included in the kit

- ddH<sub>2</sub>O for dissolving Proteinase K
- RNase A (10 mg/ ml), optional
- 1.5 ml and 2.0 ml tubes

## 7 Initial steps before starting

- Ensure that the **Proteinase K** has been prepared according to the instruction (→ “Kit components” p. 7).
- Avoid freezing and thawing of starting material.
- Invert the Reagent Plate / Reagent Strips for 3–4 times and thump it onto a table to collect the prefilled solutions at the bottom of the wells.
- Heat thermal mixer or water bath at 65 °C.
- Centrifugation steps should be carried out at room temperature.

## 8 Product specifications

### 1. Starting material:

- Food samples (max. 200 mg)

### 2. Time for isolation:

#### - Manuel steps:

- Lysis: approx. 60 minutes
- Processing after lysis: approx. 15 minutes

#### - Automated steps:

Extraction protocol InnuPure C16 / C16 <i>touch</i>	Protocol on In- nuPure C16 / C16 <i>touch</i>	Time In- nuPure C16 / C16 <i>touch</i>	Elution volumes
Ext_Lysis_200_C16_04/ External Lysis 200 µl – 05	200 µl	55 / 52 min	20–500 µl
Ext_Lysis_200_Fast_C16_04/ External Lysis 200 µl – Fast – 05	200 µl	43 / 41 min	20–500 µl

### 3. Typical yield:

- Depending on amount and quality of starting material.

## 9 Protocol: Lysis of food samples

1. Weigh up to **200 mg** of food sample and transfer it into a 2.0 ml tube. Before, cut the sample in small pieces or homogenize the sample as much as possible.
2. Add the recommended amount of **Lysis Solution CBV** (see table) and **20 µl Proteinase K** to each sample and vortex vigorously for 10 seconds. Incubate at 65 °C for approx. 60 minutes.

Food class	Example	Amount of Lysis Solution CBV to be added to the sample
Meat products	ham, salami	0.8 ml
Tinned food	fish, meat or sausages	0.8 ml
Milk products	cheese, yoghurt, chocolate	0.8 ml
Cereals	flakes, nachos, waffle, cookie, noodle	1.5 ml
Flours	wheat flour, baking mixes	1.2 ml
Instant products	Instant soups, mashed potatoes	1.0 ml

### NOTE

We recommend to use a shaking platform (thermal mixer, water bath or another rocking platform) for a continuous shaking of the sample. Alternatively, vortex the sample every 10 minutes during the incubation. No shaking will reduce the lysis efficiency.

3. Centrifuge the tube at 11,000 x g (11,000 rpm) for 10 minutes.
4. Transfer the supernatant into a new 1.5 ml reaction tube.  
If there is a floating material above the sample, pierce this film carefully with pipette and carefully remove the sample. Avoid aspiration of floating material and/or sediment.

### NOTE

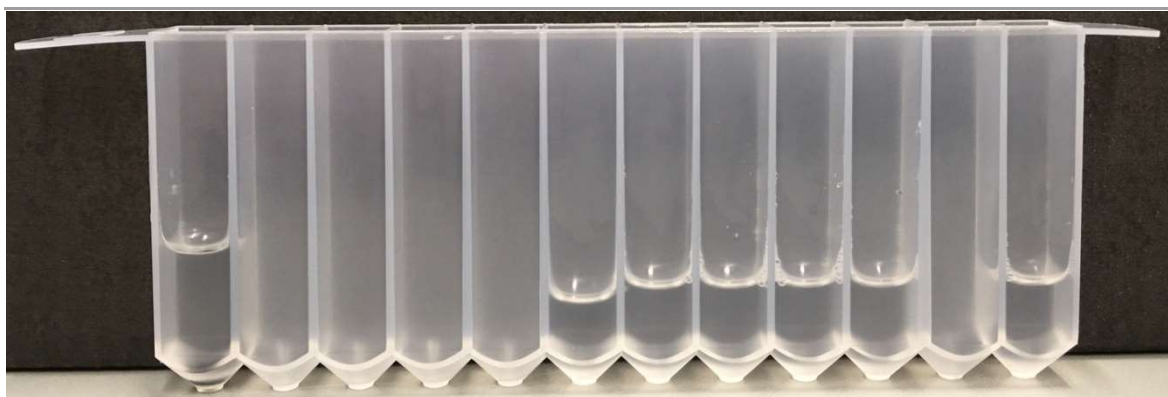
To remove RNA from the sample (if necessary) add 2  $\mu\text{l}$  of RNase A solution (10 mg/ml) to the lysed sample, vortex shortly and incubate for 5 minutes at room temperature.

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5. Check if the sample volume is at least **400  $\mu\text{l}$** . If it is lower fill up by addition of **Lysis Solution CBV**.
6. Proceed with automated extraction ( $\rightarrow$  "Preparing Reagent Plate / Strip for automated extraction", p. 10).

## 10 Preparing Reagent Plate / Strip for automated extraction

### 10.1 General filling scheme of reagent reservoir



Cavity 1:	Magnetic particles	Cavity 7:	Washing Solution
Cavity 2:	Empty	Cavity 8:	Washing Solution
Cavity 3:	Empty	Cavity 9:	Washing Solution
Cavity 4:	Empty	Cavity 10:	Washing Solution
Cavity 5:	Empty	Cavity 11:	Empty
Cavity 6:	Binding Solution	Cavity 12:	Elution Buffer

### 10.2 Unpacking of Reagent Plate or Reagent Strip

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#### NOTE

According to transport regulations Reagent Reservoirs are wrapped into plastic bags only when transported by airplane.

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Reagent Plates or Reagent Strips are delivered wrapped into plastic bags for transport protection.

Carefully open the overpack of Reagent Plates or Strips by using scissors.

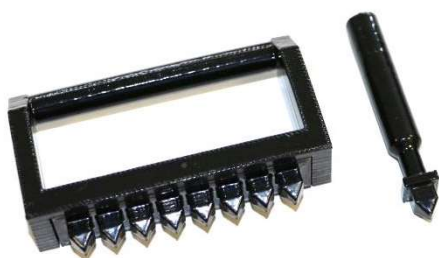
### 10.3 Piercing of sealing foil of Reagent Plate or Reagent Strip

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#### NOTE

Before using Reagent Plates or Strips the sealing foil has to be pierced manually. Always wear gloves while piercing of the foil!

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Reagent Plates or Strips are prefilled with extraction reagents and are sealed with a foil. Before use this foil has to be pierced manually, by using the piercing tools (single piercer or 8fold piercer).

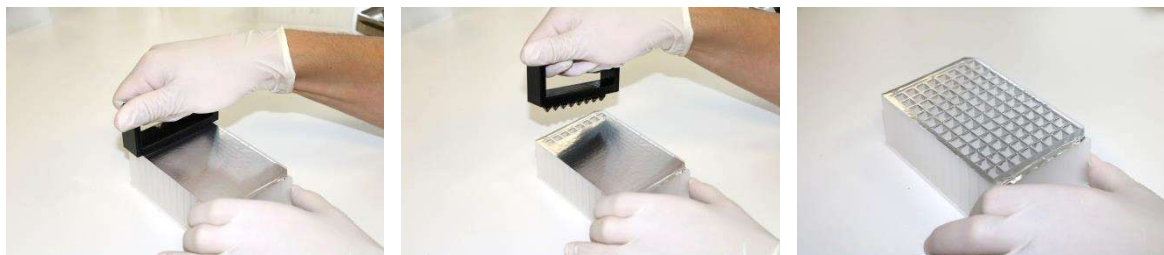
Keep the Reagent Plates or Strips in a horizontal position to avoid spilling of the reagents while piercing of the foil.

Open all cavities (one row per sample).

## Preparing Reagent Plate / Strip for automated extraction

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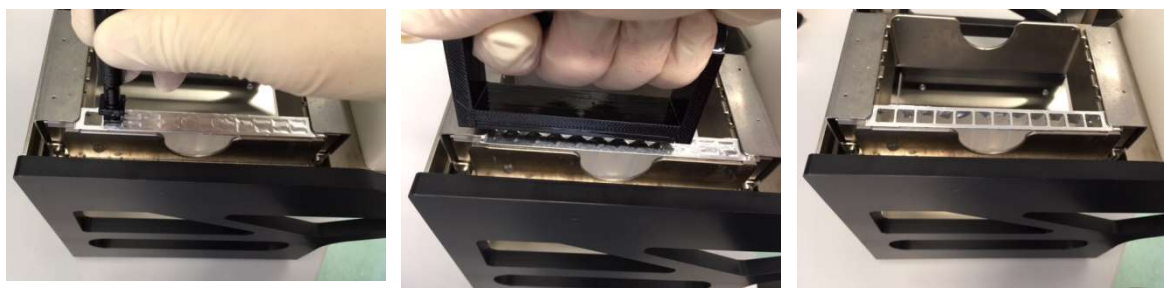
### Using 8 samples in parallel



### Using single samples



### Using Reagent Strips



### IMPORTANT

Use single or eightfold piercing tool for opening of all cavities of one row per sample!

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## 10.4 Loading the sample to InnuPure C16 / C16 touch

1. Ensure the foils of Reagent Plate or Reagent strips have been pierced (→ „Preparing Reagent Plate / Strip for automated extraction“ p. 10).
2. Transfer 400 µl of lysed sample directly into the **third cavity** of Reagent Plates or Reagent Strips.

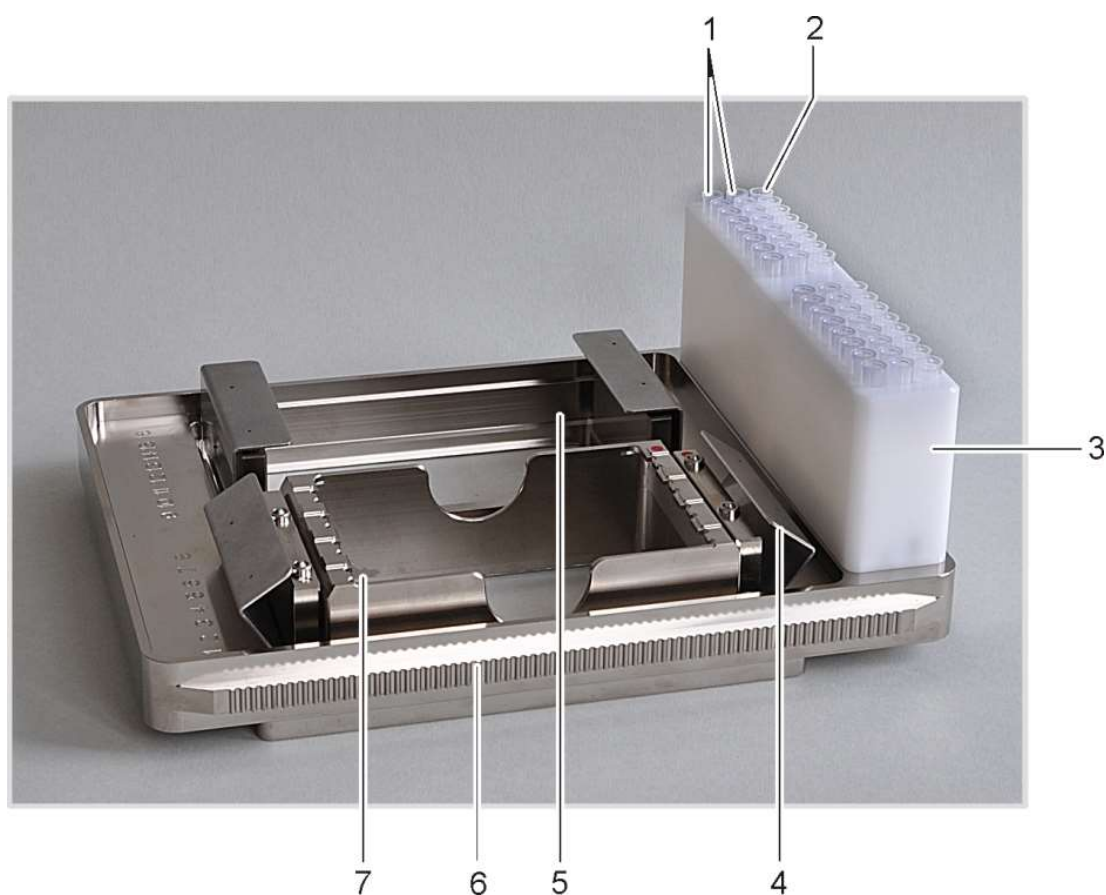
### NOTE

The sample will be processed using the InnuPure C16 / C16 touch. Please follow the instruction of chapter 11 p. 13.

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## 11 Automated extraction using InnuPure C16 / C16 touch

### 11.1 Sample tray of InnuPure C16 / C16 touch



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**No. 1:** Filter tips

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**No. 2:** Elution vessels for purified samples

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**No. 3:** Tip block

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**No. 4:** Holding-down clamp

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**No. 5:** Sample block for Reagent Plates or adapter for Reagent Strips

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**No. 6:** Serrated guide rail (C16 touch: non-serrated)

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**No. 7:** Adapter for Reagent Strips

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## 11.2 Preparing sample tray of InnuPure C16 / C16 touch

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### NOTE

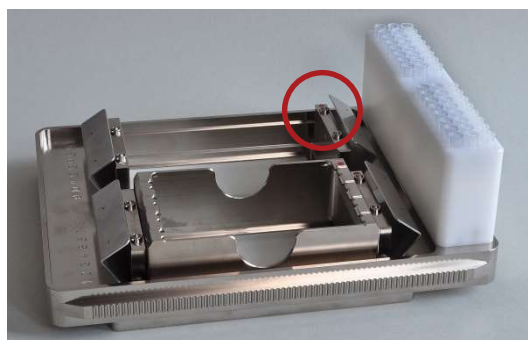
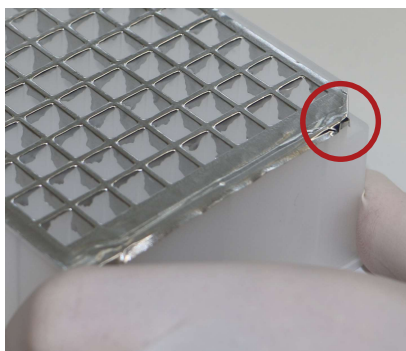
The needed number of Reagent Strips or Reagent Plates is depending on the number of samples, which have to be processed. Don't use more strips as number of samples!

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1. Place the InnuPure C16 / C16 *touch* sample tray into the priming station and fold the holding-down clamp at the sample tray upwards!
2. Place the Reagent Plate or an adapter with Reagent Strips into the holder of the sample tray. Using Reagent Plates, the notched corner of the Reagent Plate has to align with the colored dot at the holder. Using adapters and Reagent Strips, the colored dot of the adapter has to align with the colored dot at the holder and Reagent Strips have to be inserted in a way that the "AJ" labels are arranged at the side of the adapter which is more distant from the tip block.

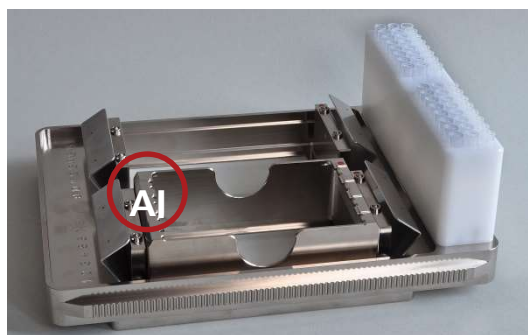
### Reagent Plate

The notched corners of the Reagent Plate must point to the colored dot on the holder.



### Reagent Strips

Place the Reagent Strips into the adapter. The long tab marked with the label "AJ" must point to the side of the adapter which is more distant from the tip block.





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**ATTENTION**

Both holders have to be equipped with a Reagent Plate or Reagent Strips. If applicable use an empty or dummy plate for the respective holder.

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3. Fold down the holding-down clamp to prevent the Reagent Plates and Reagent Strips to be pulled out of the holder during the extraction process.
  4. For each extracted sample place two filter tips in the smaller drill holes of the tip block.
  5. Place the Elution Tubes into the wider drill hole at the edge of the tip block. Empty sample positions do not need to be filled.
- 

**NOTE**

Especially with the Reagent Strips make sure that for every Reagent Strip the tips and the elution vessel are in the corresponding positions in the tip block!

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**ATTENTION**

It is possible to select between two different elution vessels! For small elution volumes up to 200 µl use Elution Strips (0.2 ml). For high elution volumes up to 500 µl use Elution Tubes (0.65 ml) with corresponding Elution Caps (Stripes).

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### 11.3 Starting the InnuPure C16

1. Switch on the InnuPure C16 and wait for the device initialization to complete, which is signaled by a beeping sound.
2. Move the loaded sample tray with the Reagent Strips or Reagent Plates forward into the sample tray adapter on the front of the InnuPure C16. The serrated rails at the side of the sample tray must protrude into the grooves of the adapter. After pressing lightly against the tip block the sample tray is automatically pulled into the device.



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#### ATTENTION

Immediately let go of the sample tray it is being pulled in. Otherwise there is a risk of your hand being crushed.

---

3. After pressing [Select Protocol] choose an appropriate extraction protocol on InnuPure C16 and press [Start]:

Extraction procedure	Protocol on InnuPure C16
Standard (maximum yield, approx. 55 minutes)	Ext_Lysis_200_C16_04
Fast (time-optimized, approx. 43 minutes)	Ext_Lysis_200_Fast_C16_04

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- 4.

5. Enter the recommended **elution volume** of **150–200 µl** and press [OK].
- 

**NOTE**

It is possible to adjust the volume values from 20 µl to 500 µl.

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6. If needed, choose log file and enter sample ID's, press [OK] or [CANCEL].
- 

**NOTE**

It is possible to enter sample ID's and to create a run log file. Find more detailed information how to start an extraction protocol using InnuPure C16 on page 37 of the user manual ("6.3.5 Using the sample set-up tool")!

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7. After completion of the protocol press [NEXT] and the sample tray is then automatically moved out of the device.
- 

**NOTE**

The chosen protocol is performed by the device and after the protocol is finished, the tray with the purified samples will be moved out after pressing [NEXT] and the message 'Program finished' is shown on the screen of the device!

---

8. Remove the sample tray from the adapter of the InnuPure C16 and place it back into the priming station.
  9. After finishing the extraction protocol, the Elution Tubes contain the extracted DNA or RNA. Close the lids and store the DNA under proper conditions.
- 

**NOTE**

Store DNA and RNA under adequate conditions. We recommend storing the extracted DNA at -22 °C to -18 °C!

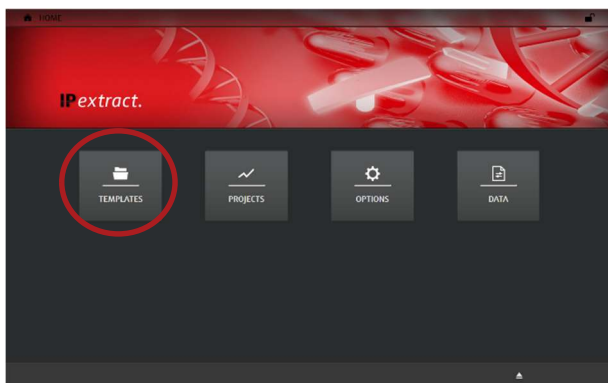
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## 11.4 Starting the InnuPure C16 touch

### NOTE

The following instructions describe the necessary steps for the start of the InnuPure C16 touch. For further features and data entry (e.g. opening templates, entering sample setups, saving projects) refer to the manual of the InnuPure C16 touch.

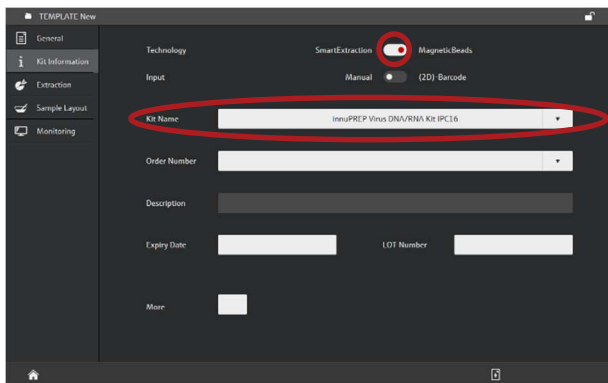
1. Switch on the InnuPure C16 touch and the tablet computer. Wait until the home screen of IPextract is displayed on the tablet screen.



### NOTE

Home screen of IPextract

2. Choose [TEMPLATES] → [New Template] → [Kit-based].
3. Enter optional information in the tab "General".
4. Choose the tab "Kit Information" and switch the "Technology" to "MagneticBeads".
5. Choose your desired kit from "Kit Name".



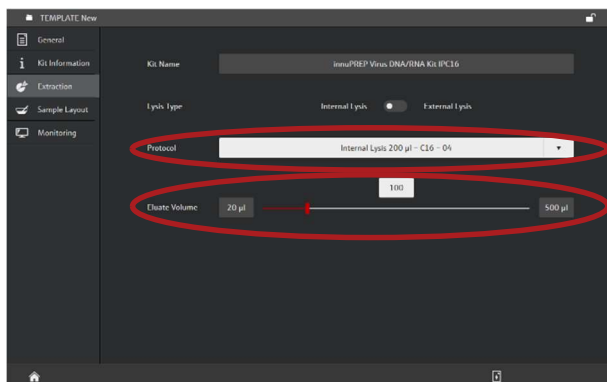
### NOTE

"Kit Information" tab

6. Enter optional information in the tab "Kit Information"
7. Choose the tab "Extraction" and choose the desired "Protocol"

Extraction procedure	Protocol on InnuPure C16 <i>touch</i>
<b>Standard</b> (maximum yield, approx. 52 minutes)	External Lysis 200 µl - 05
<b>Fast</b> (time-optimized, approx. 41 minutes)	External Lysis 200 µl - Fast - 05

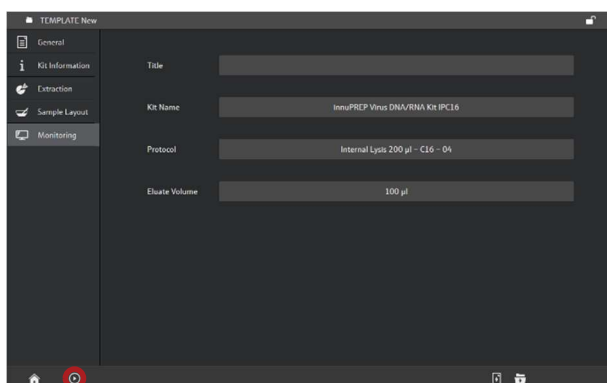
8. Adjust your desired "Eluate Volume" using the slider or the text field.



**NOTE**  
"Extraction" tab

The recommended elution volume is 150–200 µl.

9. Choose the tab "Monitoring" and start the protocol by tapping the start button.



**NOTE**  
"Monitoring" tab

10. Follow the instructions displayed on the tablet screen.
11. Completion of the protocol is indicated by a message on the tablet screen. Follow the instructions on the screen to remove the sample tray from the device.
12. The Elution Tubes contain the extracted DNA or RNA. Close the lids and store the DNA under proper conditions.

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### NOTE

Store the DNA and RNA under adequate conditions. We recommend storing the extracted DNA at -22 °C to -18 °C!

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## 12 Troubleshooting

Problem / probable cause	Comments and suggestions
<b>Low amount of extracted DNA</b>	
Insufficient lysis of starting material	Ensure to use the required volume of <b>Proteinase K</b> and amount of <b>Lysis solution CBV</b> .
Eluate volume too high	Decrease the eluate volume. The suggested eluate volume is 150–200 µl. Please note that lowering the eluate volume will not necessarily increase the yield proportionally!
Inadequate extraction	Inhibiting substances in starting material. Please use the kit only for samples that match the requirements declared in "Product specifications".

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